2004 Vol. 6, No. 18 3087-3089

Citrinadin A, a Novel Pentacyclic Alkaloid from Marine-Derived Fungus *Penicillium citrinum*

Masashi Tsuda,[†] Yuu Kasai,[†] Kazusei Komatsu,[†] Teruo Sone,[‡] Michiko Tanaka,[‡] Yuzuru Mikami,[§] and Jun'ichi Kobayashi*,[†]

Graduate School of Pharmaceutical Sciences, Hokkaido University, Sapporo 060-0812, Japan, Graduate School of Agriculture, Hokkaido University, Sapporo 060-8589, Japan, and Research Center for Pathogenic Fungi and Microbial Toxicoses, Chiba University, Chiba 260-0856, Japan jkobay@pharm.hokudai.ac.jp

Received June 11, 2004

ABSTRACT

citrinadin A (1)

A novel pentacyclic alkaloid, citrinadin A (1), was isolated from the cultured broth of the fungus *Penicillium citrinum*, which was separated from a marine red alga, and the structure was elucidated by spectroscopic data. The relative stereochemistry of the pentacyclic core was assigned on the basis of NOESY data and ¹H–¹H coupling constants, and the presence of an *N,N*-dimethyl-L-valine residue in 1 was determined by chiral HPLC analysis of the hydrolysate.

Marine-derived fungi have proven to be a rich source of structurally unique and biologically active secondary metabolites. In our search for new metabolites from marine-derived fungi, a novel pentacyclic spiroindolinone alkaloid, citrinadin A (1), with an N,N-dimethylvaline residue and an α,β -epoxy carbonyl unit, was isolated from the cultured broth of a fungus *Penicillium citrinum*, which was separated from a marine red alga. In this paper, we describe the isolation and structure elucidation of 1.

The fungus *Penicillium citrinum* (strain N-059) was separated from the red alga *Actinotrichia fragilis* collected

at Hedo Cape, Okinawa Island, and grown in PYG liquid medium containing seawater for 14 days at 25 °C. The mycerium (435 g) of the culture broth (12 L) was extracted with MeOH. The extract was partitioned between hexane and 90% aqueous MeOH, and the MeOH-soluble portion was extracted with *n*-BuOH. The *n*-BuOH-soluble portions were subjected to LH-20 and SiO₂ column chromatographies to afford the bis-salt of citrinadin A (1, 6.5 mg, 0.0015%, wet weight).

The bis-salt of citrinadin A³ {1, $[\alpha]^{19}_D$ -17° (c 0.4, MeOH)} showed the pseudomolecular ion peak at m/z 625 in the FABMS, and the molecular formula was revealed to be $C_{35}H_{52}O_6N_4$ by HRFABMS [m/z 625.3964, (M + H)⁺,

^{*} To whom correspondence should be addressed. Phone: +81-11-706-4985. Fax: +81-11-706-4989.

[†] Graduate School of Pharmaceutical Sciences, Hokkaido University.

[‡] Graduate School of Agriculture, Hokkaido University.

[§] Chiba University.

^{(1) (}a) Blunt, J. W.; Copp, B. R.; Munro, M. H. G.; Northcote, P. T.; Princep, P. R. *Nat. Prod. Rep.* **2004**, *21*, 1–49. (b) Blunt, J. W.; Copp, B. R.; Munro, M. H. G.; Northcote, P. T.; Princep, P. R. *Nat. Prod. Rep.* **2003**, 20, 1–48. (c) Faulkner, D. J. *Nat. Prod. Rep.* **2002**, *19*, 1–48 and references therein.

^{(2) (}a) Komatsu, K.; Shigemori, H.; Kobayashi, J. J. Org. Chem. **2001**, 66, 6189–6192. (b) Tsuda, M.; Mugishima, T.; Komatsu, K.; Sone, T.; Tanaka, M.; Mikami, Y.; Shiro, M.; Hirai, M.; Ohizumi, Y.; Kobayashi, J. Tetrahedron **2003**, 59, 3227–3230.

Table 1. ¹H and ¹³C NMR Data of the Bis-Salt of Citrinadin A (1) in CDCl₃

position	δ_{C}	m		$\delta_{ m H}$	m, Hz
1				9.64	s
2	184.89	S			
3	60.42	s			
3a	134.52	s			
4	133.30	d		7.66	d, 7.4
5	122.49	d		7.19	t, 7.7
6	127.64	d		7.78	d, 8.0
7	117.52	S			
7a	142.67	S			
8	41.52	t		2.20^{a}	S
9	67.90	S			
10	51.31	t	(a)	3.26	d, 11.4
			(β)	3.81	m
11			4 /	11.27	brs
12	56.12	d		3.69	m
13	32.70	t	(a)	1.88	brd, 16.0
			(β)	3.47	m
14	68.92	d	4 /	5.43	brs
15	33.34	t	(a)	2.00	brd, 14.6
			(β)	3.13	brt, 13.8
16	47.48	d	4.7	4.07	m
17	31.35	t	(a)	1.80	brd, 12.0
			(β)	2.17	brt, 12.0
18	82.13	s	4.7		,
18-OH				5.38	brs
19	51.00	s			
20	194.84	s			
21	64.06	d		4.05	s
22	61.72	s		1.00	
23	24.31	q		1.60^{b}	S
24	18.62	q		1.37^{b}	S
25	10.02	4		2.75	br
26	30.57	q		2.50^{b}	S
27	15.21	q		1.59^{b}	d, 6.4
28	21.93	q		1.02^{b}	s s
29	27.96	q q		1.37^{b}	S
1'	166.72	s S		1.07	5
2'	71.38	d		3.77	d, 6.7
~ 3′	71.00	u		12.68	brs
4′	28.35	d		2.40	m
5'	19.47			1.07^{b}	d, 6.5
5 6′	21.37	q		1.07^{b}	d, 6.5
0 7′	38.34	q		2.99^{b}	u, o.s brs
, 8'	43.14	q q		2.99^{b} 2.94^{b}	brs
Q'					

-0.1 mmu]. The IR spectrum suggested the presence of OH/NH (3401 cm $^{-1}$) and carbonyl group(s) (1710 and 1671 cm $^{-1}$). The UV absorption at 335 nm (ϵ 3100) was attributed to a conjugated benzenoid chromophore. The ^{13}C NMR (Table 1) spectrum disclosed the existence of three carbonyls (δ_C 194.84, 184.89, and 166.72), three sp 2 quaternary carbons (δ_C 142.64, 134.52, and 117.52), three sp 2 methines (δ_C 133.30, 127.64, and 122.49), five sp 3 quaternary carbons (δ_C

82.13, 67.90, 61.72, 60.42, and 51.00), six sp³ methines ($\delta_{\rm C}$ 71.38, 68.92, 64.06, 56.12, 47.48, and 28.35), five sp³ methylene ($\delta_{\rm C}$ 51.31, 41.52, 33.34, 32.70, and 31.35), and 10 methyls ($\delta_{\rm C}$ 43.14, 38.34, 30.57, 27.96, 24.31, 21.93, 21.37, 19.47, 18.62, and 15.21). Because six out of 12 unsaturations were accounted for, **1** was inferred to contain six rings.

The 1 H NMR (Table 1) spectrum included five D₂O-exchangeable proton signals (δ_H 12.68, 11.27, 9.64, 5.38, and 2.75) due to NH and/or OH groups. One (δ_H 11.27, NH-11) of two low-field D₂O-exchangeable resonances showed cross-peaks to H-10 β (δ_H 3.81), H-12 (δ_H 3.69), and H-16 (δ_H 4.07) in the 1 H COSY spectrum, whereas the other (δ_H 12.68, NH-1) correlated with a methine (δ_H 3.77, H-2') and two *N*-methyl signals (δ_H 2.99, H₃-7'; δ_H 2.94, H₃-8') in the 1 H- 1 H COSY and the TOCSY spectra. These observations suggested that the two resonances were attributed to ammonium NH signals at N-11 and N-3'. The 1 H- 1 H COSY, TOCSY, and HSQC spectra revealed four connectivities from C-4 to C-6, from C-10 to C-17 and C-27 through N-11, from NH-25 to C-26, and from C-4' to C-8' through N-3' and C-2' (Figure 1). The existence of a 1,2,3-

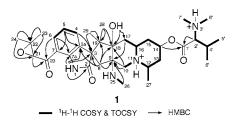


Figure 1. Selected 2D NMR correlations for citrinadin A (1).

trisubstituted benzene ring was indicated by HMBC correlations for H-4 ($\delta_{\rm H}$ 7.66)/C-3, H-4/C-7a ($\delta_{\rm C}$ 142.67), H-5 ($\delta_{\rm H}$ 7.19)/C-3a ($\delta_{\rm C}$ 134.52), H-5/C-7 ($\delta_{\rm C}$ 117.52), and H-6 ($\delta_{\rm H}$ 7.78)/C-7a. HMBC correlations were observed for NH-1 ($\delta_{\rm H}$ 9.64)/C-2 ($\delta_{\rm C}$ 184.89), NH-1/C-3 ($\delta_{\rm C}$ 60.42), NH-1/C-3a, NH-1/C-7a, and H-4/C-3, suggesting that 1 possessed an indolinone ring (C-1-C-7a). The presence of 2,3-epoxy-3methyl-1-oxobutyl side chain (C-20-C-24) at C-7 was deduced from HMBC correlations for H-7/C-20 ($\delta_{\rm C}$ 194.84), H-21 ($\delta_{\rm H}$ 4.05)/C-20, H₃-23 ($\delta_{\rm H}$ 1.60)/C-21 ($\delta_{\rm C}$ 64.06), H₃-23/C-22 (δ_C 61.72), H_3-24 (δ_H 1.37)/C-21, and $H_3-24/C-22$. ¹H NMR data of this C₅ unit in **1** were similar to those of the corresponding part of hopeyhopin.⁴ Long-range H-C couplings for H-10 α ($\delta_{\rm H}$ 3.26)/C-9 ($\delta_{\rm C}$ 67.90), H-10 α /C-18 $(\delta_{\rm C} 82.13)$, and H-17 β $(\delta_{\rm H} 2.17)$ /C-9 suggested the presence of a quinolizidine ring system (N-11 and C-9-C-18). HMBC correlations for a D₂O-exchangeable proton (OH-18) at $\delta_{\rm H}$ 5.38 to C-17 and C-18 indicated that a hydroxyl group was attached to C-18. It was revealed that an N-methylamino group (N-25-C-26) was connected to C-9 since an HMBC

3088 Org. Lett., Vol. 6, No. 18, 2004

⁽³⁾ **Bis-salt of citrinadin A (1):** colorless oil; UV (MeOH) λ_{max} 335 (ϵ 3100), 265 (sh.), 249 (9600), 230 (sh.), and 224 (9000); IR (neat) ν_{max} 3401, 2930, 1710, 1671, and 1602 cm $^{-1}$; FABMS m/z 625 (M + H) $^{+}$; HRFABMS m/z 625.3964 [(M + H) $^{+}$, calcd for C₃₅H₅₃O₆N₄, 625.3965].

⁽⁴⁾ Domínguez, X. A.; Cano, G.; Luna, I.; Dieck, A. *Phytochemistry* **1977**, *16*, 1096.

correlation was observed for H_3 -26 (δ_H 2.50)/C-9. In the HMBC spectrum, a singlet proton signal at $\delta_{\rm H}$ 2.20 (2H, H_2 -8) showed correlations to C-9, C-18, and C-19 (δ_C 51.00), and both of two singlet methyl signals at $\delta_{\rm H}$ 1.02 (H₃-28) and 1.37 (H₃-29) were correlated to C-18 and C-19, suggesting the presence of a cyclopenta[b]quinolizidine moiety (N-11, C-3, and C-8-C-19). HMBC correlations for H_2 -8/ C-3, H₂-8/C-3a, and H₃-29/C-3 indicated that the cyclopenta-[b]quinolizidine moiety and the indolinone ring were connected to each other through the spiro carbon (C-3). The presence of an N,N-dimethylvaline residue was deduced from the HMBC correlation for H-2'/C-1' ($\delta_{\rm C}$ 166.72). The $^{13}{\rm C}$ chemical shift at C-1' were close to those of the N,Ndimethylvaline ester in 14-(N,N-dimethyl-L-valyloxy)paspalinine⁵ rather than those of the N.N-dimethylvalinamide terminus in dolastatin 10,6 indicating that 1 possessed an N,Ndimethylvaline ester. The relatively low-field chemical shift of H-14 ($\delta_{\rm H}$ 5.43) suggested that the N,N-dimethylvaline residue was attached to C-14 via an ester linkage.⁷ [This was supported by high-field shift of H-14 by hydrolysis of the N,N-dimethylvalyl ester (vide infra).] Therefore, the gross structure of citrinadin A was concluded to be 1.

The relative stereochemistry of the pentacyclic core of citrinadin A (1) was elucidated on the basis of ROESY data and ¹H-¹H coupling constants (Figure 2). ROESY correla-

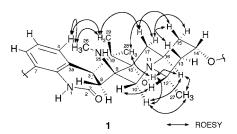


Figure 2. Relative stereochemistry for the pentacylic ring core of citrinadin A (1).

tions for H-4/H₃-26 and H-4/H₃-29 indicated that one (C-29) of two methyl groups at C-19, the C-3—C-3a bond, and the methylamino group (C-25—C-26) were β -oriented. Axial orientations for the β -protons at C-13, C-15, and C-17 were implied from ROESY correlations for H-13 β /H-15 β , H-15 β /H-17 β , and H-17 β /H₃-29 and proton signal patterns for H-15 β (brt, J=13.8 Hz) and H-17 β (brt, J=12.0 Hz). On the other hand, ROESY correlations for H-10 α /H₃-27, H-10 α /OH-18, H-16/OH-18, H-16/H₃-27, and OH-18/H₃-28 suggested α -axial orientations for H-10 α , H-16, OH-18, and

 H_3 -27. Because ROESY correlations were observed for H-10 β /H-12, H-15 α /H-17 α , and H-17 α /H₃-28, H-10b, H-12, H-15a, and H-17a were considered to have equatorial orientations. The broad singlet pattern of H-14 suggested that the oxygen atom at C-14 was α-axially oriented. Therefore, the relative configuration of the cyclopenta[b]quinolizidine moiety in **1** was elucidated to be anti/syn/anti and chair forms for two six-membered rings. On the other hand, it was difficult to elucidate unambiguously the relative configuration of the epoxide ring at C-21–C-22.

The absolute configuration at C-2' in the N,N-dimethylvaline residue was determined on the basis of chiral HPLC analysis of the hydrolysate of citrinadin A (1). Hydrolysis of 1 with 1 N aqueous HCl afforded 1 mol of N,N-dimethylvaline. The N,N-dimethylvaline residue was identified as L-form by chiral HPLC analysis using authentic D-and L-N,N-dimethylvaline.

Citrinadin A (1) is a novel pentacyclic spiroindolinone alkaloid with an N,N-dimethylvaline ester and an α,β -epoxy carbonyl unit. There are only two prior reports of the natural occurrence of the N,N-dimethylvaline residue describeed in 14-(N,N-dimethyl-L-valyloxy)paspaline⁵ and dolastatins,⁶ which were isolated from fungi and sea hares, respectively. Although several spiroindolinone alkaloids such as brevianamides,⁹ paraherquamides,¹⁰ marcfortines,¹¹ sclerotamide,¹² and asperparalins¹³ have been isolated from fungi of the genera *Penicillium* or *Aspergillus*, the pentacylcic skeleton such as 1 is unique. Citrinadin A (1) exhibited cytotoxicity against murine leukemia L1210 and human epidermoid carcinoma KB cells (IC₅₀ 6.2 and 10 μ g/mL, respectively).

Acknowledgment. We thank M. Kiuchi, Center for Instrumental Analysis, Hokkaido University, for FABMS measurements, and M. Iha, Southproduct Co. Ltd., for his help with red alga collection. This work was partly supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

Supporting Information Available: Experimental procedures and spectral data of **1**. This material is available free of charge via the Internet at http://pubs.acs.org.

OL048900Y

Org. Lett., Vol. 6, No. 18, 2004

⁽⁵⁾ Staub, G. M.; Gloer, K. B.; Gloer, J. B. Tetrahedron Lett. 1993, 34, 2569–2572.

⁽⁶⁾ Pettit, G. R.; Kamano, Y.; Herald, C. L.; Tuinman, A. A.; Boettner, F. E.; Kizu, H.; Schmidt, J. M.; Baczynsky, J. L.; Tomer, K. B.; Bontems, R. J. J. Am. Chem. Soc. 1987, 109, 6883–6885.

⁽⁷⁾ The corresponding proton of the 2-dihydro form of myrtine resonated at $\delta_{\rm H}$ 3.99 in CDCl₃. Solsse, P.; Hootelé, C. *Tetrahedron* **1981**, *37*, 4287–4292.

⁽⁸⁾ Bowman, R. E.; Stroud, H. H. *J. Chem. Soc.* **1950**, 1342–1345. (9) (a) Birch, A. J.; Wright, J. J. *J. Chem. Soc.* **D 1969**, 644–645. (

^{(9) (}a) Birch, A. J.; Wright, J. J. *J. Chem. Soc. D* **1969**, 644–645. (b) Birth, A. J.; Wright, J. J. *Tetrahedron* **1970**, 26, 2329–2344. (c) Birch, A. J.; Russell, R. A. *Tetrahedron* **1972**, 28, 2999–3008.

^{(10) (}a) Yamazaki, M.; Okuyama, E.; Kobayashi, M.; Inoue, H. *Tetrahedron Lett.* **1981**, 22, 135–136. (b) Ondeyka, J. G.; Goegelman, R. T.; Schaeffer, J. M.; Keleman, L.; Zitano, L. *J. Antibiot.* **1990**, 43, 1375–1379. (c) Blanchflower, S. E.; Banks, R. M.; Everett, J. R.; Manger, B. R.; Reading, C. *J. Antibiot.* **1991**, 44, 492–497. (d) Blanchflower, S. E.; Banks, R. M.; Everett, J. R.; Reading, C. *J. Antibiot.* **1993**, 46, 1355–1363.

^{(11) (}a) Polonsky, J.; Merrien, M.-A.; Prangé, T.; Pascard, C.; Moreau, S. *J. Chem. Soc., Chem. Commun.* **1980**, 601–602. (b) Prangé, T.; Billion, M.-A.; Vuilhorgne, Pascard, C.; Polonsky, J. *Tetrahedron Lett.* **1981**, 22, 1977–1980.

⁽¹²⁾ Whyte, A. C.; Gloer, J. B. J. Nat. Prod. 1996, 59, 1093-1095.

^{(13) (}a) Hayashi, H.; Nishimoto, Y.; Nozaki, H. Tetrahedron Lett. 1997, 38, 5655–5658. (b) Hayashi, H.; Nishimoto, Y.; Akiyama, K.; Nozaki, H. Biosci. Biotechnol. Biochem. 2000, 64, 111–115.